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EVALUATION OF STABILITY AND CRESTAL BONE CHANGES AROUND IMPLANTS PLACED AT CRESTAL VERSUS SUBCRESTAL LEVEL IN CONTROLLED TYPE TWO DIABETIC PATIENTS (RANDOMIZED CLINICAL TRIAL)

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ABSTRACT

INTRODUCTION:Peri-implant bone level preservationis the key to maintainingperi-implant soft tissue and stability.Many studies have discussed the role of placing dental implants at various depths on crestal bone loss, but they were debatable.
OBJECTIVES: To compare clinically and radiographically the effect of placing implants atcrestal versus subcrestal levels on the crestal bone lossand stability evaluation in controlled type 2diabetic patients.
MATERIALS AND METHODS: 22 controlleddiabetic patients were randomized accordingto placement depth (group I: 11 implants were placed equicrestally) (group II: 11implants were placed 1mm subcrestally). Stability of implants, clinical and radiographical assessment were done for both groups.
RESULTS: 19 implants were included in our study (10 crestal and 9 subcrestal). There were no statistically significant differences in bleeding index and probing depth between the 2 groups while plaque index was greateramong subcrestal group. Crestal bone loss mesiodistally was significantly higherin crestal group, whilethere was no significant difference buccolingually between crestal and subcrestal groups. Stabilityvalues weresignificantly greater in subcrestal group at loading time.
CONCLUSION: Sub-crestal implant placement is preferable for controlled type 2 diabetic patients as it decreases the probability of implant thread exposureprovided that careful oral hygiene care is followed and regular periodic checks to maintain peri-implantsoft tissue and dental bone health.
KEYWORDS: Crestal, Subcrestal, Stability, Type 2 diabetes mellitus.
RUNNING TITLE: Crestal Bone Loss And Stability Evaluation In Type 2 Diabetic Patients.

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INTRODUCTION

Dental implants have become a successful alternative in the treatment of partially or completely edentulous patients for restoring function and esthetics (1). It is considered that crestal loss up to 1.5 mmis a normal findingthrough the firstyear post the placement of the implant and then followed by less than 0.2 mm annually (2). A lot of studies have focused on the preservation of peri-implant bone level and soft tissue which are considered crucial factors for long-term implant stability and also for maintaining esthetics (3).

There are a lot of local factors affecting the stability of marginal bone around the dental implant and subsequently implant survival such as biologic width formation, bone density, surgical trauma, peri-implantitis, gingival biotype, stress concentration at crestal bone level, and location of the implant-abutment connection. On the other hand, many systemic conditions may affect implant survival such as (diabetes, cardiovascular diseases, elderly populations, bisphosphonate therapy, and bleeding disorders) (4).

Diabetes Mellitus is considered one of the most common metabolic-related endocrine diseases that is characterized by hyperglycemia as a result of a deficiency in insulin secretion (type 1) or insulin resistance (type 2). Type 2 diabetes represents about 90%-95% of all those individuals who aresuffering from Diabetes Mellitus (5). Implant therapy in patients with type 2 diabetes mellitus (T2DM) was considered a critical treatment option because of the increased risk of infections and delayed healing due to microangiopathy. Also, hyperglycemia may impair implant osteointegration and increase marginal bone loss by suppressing osteoblastic proliferation and differentiation (6). However, a lot of recent studies have suggested that dental implants can successfully survive functionally and esthetically under optimal glycemic control (7).

Glycosylated hemoglobin is considered a diagnostic tool for T2DM and for detecting blood sugar regulation over a period of 2 to 3 months (8). It is approved that an HbA1C level of ≤ 7% is the optimal glycemic control level in the diabetic patients while for uncontrolled patients it is more than 7% (5).

The location of the implant-abutment interface plays a crucial role in the remodeling of the marginal bone (9). Rough implant surface exposure leads to accumulation of greater amounts of plaque in this area which results in mucositis and peri-implantitis so Welander and Madani*et al* suggested that placing an implant below the bone crest is beneficial (10,11). In contrast, other studies declared that there is much less loss in the peri-implant crestal bone that occurswith the implant placement at the bone crest level thansubcrestally. (12,13).

Several clinical and radiographic studies have focused on marginal bone reduction incase of implant placement at crestal and subcrestal levels among non-diabetic patients (14,15), but there were few and controversial data carried on diabetic patients.

Several studies assessed the amount of crestal bone loss with conventional peri apical radiography which measures only mesial and distal aspects of the implant (16). However, it can`t measure the buccal or lingual aspects,despite the fact of buccal crestal bone assessmentimportance because it influences the location of mucosal marginal level and subsequently affects the esthetic outcome of the restoration(17) so, using cone beam computed tomography (CBCT) as a 3-dimensionalx-ray to assess buccolingual and mesiodistal crestal bone loss is of great importance so, we aim to compare clinically and radiographically the effect of the dental implant placement at the crestal and subcrestal levelson the crestal bone loss and the stability amongT2DM patients.

The null hypothesis is that there is no difference in the loss occurring in the crestal bone around the placed implants at either the crestal or subcrestal levels among T2DM patients.

MATERIALS AND METHODS

Ethical Approval

This study was a clinical trial that is conducted among patientschosen from the Oral and Maxillofacial Surgery Department's outpatient clinic at Alexandria University's Faculty of Dentistry and reported according to modified CONSORT guidelines (18). The Alexandria University Faculty of Dentistry's Ethical Committee validated the research protocol (IRB number: 00010556-IORG 0008839). This study has been registered and granted on ClinicalTrials.gov with registration number: (NCT05125445) All of the involved patients matched with the inclusion criteria and then signed informed consent before going on surgical operations. The research was carried out in compliance with the Helsinki Declaration for Experimentation on Human subjects (19).

Study design

Our study is considered a randomized controlled clinical trial investigation as the involved Patients were chosen randomly from individuals present in the outpatientclinic, at the Faculty of Dentistry, Alexandria University, who met the inclusion criteria. All patients were evaluated clinically and radiographically.

Study setting

Oral and Maxillofacial Department, at Faculty of Dentistry in Alexandria University.

Sample size estimation

The sample size was estimated based on assuming a confidence level= 95% and study power= 80%. The mean ± SD loss in the marginal bone subsequently after succeeding 6 months when implants were placed 1 mm subcrestally = 0.68 ± 0.39. When the implants were positioned at the bone crest, the analyzed marginal bone mean ± SD loss= 0.15 ± 0.37. The calculated sample size was considered to be 10 implants per group. This will be increased to 11 to make up for the loss to follow-up. The total sample size= number of groups × number per group= 2× 11= 22.

Inclusion criteria (20)

Patients involved in the study were those with Type 2 controlled Diabetes MellitusHBA1C ≤ 7 who were having healthy periodontium missing at least one tooth from the lower jaw with adequate height and width of the alveolar ridge and

sufficient inter-occlusal distance that accepts any future restoration, were included in the study.

Exclusion criteria (20)

Smokers, patients doing para functional habits or having bad oral hygiene, Pregnant women, and individuals suffering from any local or systemic diseases that have the probability to compromise the surgical procedurehealingor osteointegration process, were all excluded from the study.

Methods

A. Presurgical Phase

- The following data were collected from the patientsincluding name, gender,age, medical and dental history. All patients were subjected to HbA1c monitoring. Only patients who met the eligibility criteria of glycosylated hemoglobin (HbA1C) levels ≤ 7% were included in this study. Clinical evaluation of all teeth and oral tissues was done.
- CBCT images were acquired with J.Morita R100 cone beam 3D imaging systemand was used to measure the available bone, ensure the implant's right size selection for optimal support and the precision of the implant placement in the bonealso,their relation with the adjacent structures, and to evaluate the bone condition.

B. Surgical Procedure (4) Fig (1,2)

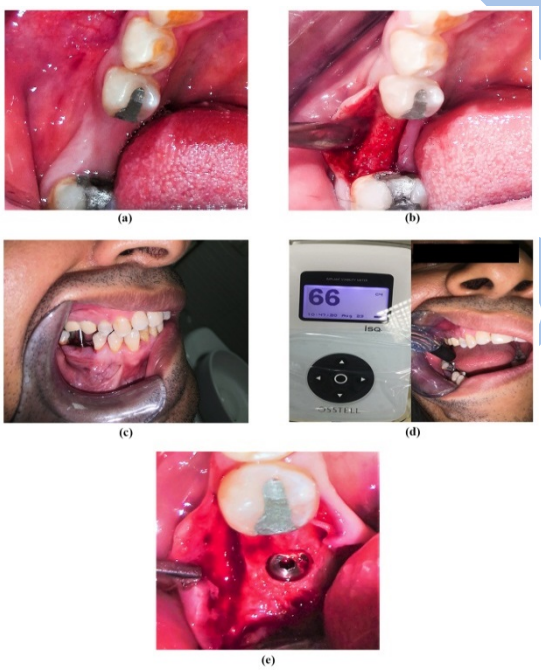


Figure 1: Surgical stage (crestal group)

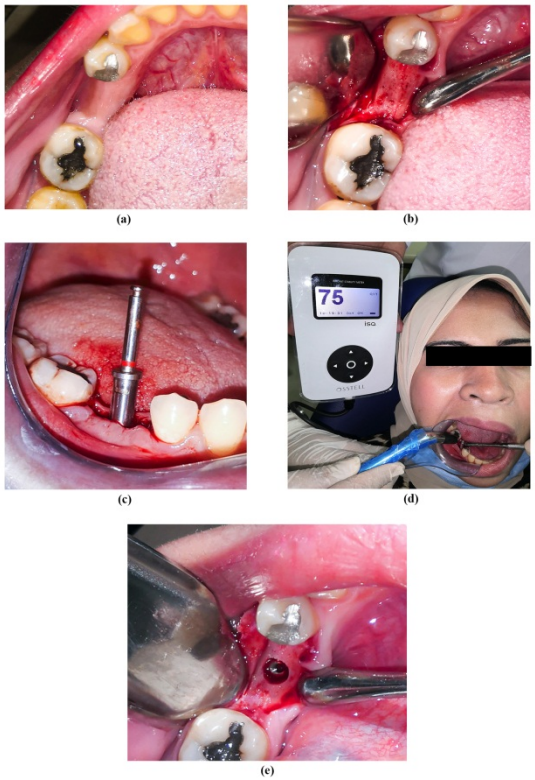


Figure 2: Surgical stage (subcrestal group)

- Systemic antibiotic prophylaxis(amoxicillin+clavulanate,Augment in, GlaxoSmithKline, Middlesex, UK) was given to patients 1 hour prior to surgery.
- All patients were informed to rinse their mouth for about 2 minutes using chlorhexidine antiseptic mouthwash before the surgery.
- Application of local anesthesia was done using 4% articaine (1:100,000 adrenaline).
- Mid crestal incision was done and an envelope flap with full thickness was raised to expose the alveolar crest.
- When required, ridge flattening was done
- The implant site was well prepared as stated by the manufacturer's instructions and performed under sterile copious irrigation. IS-II active implants, NeoBiotec were used with different lengths (8.5,10,11.5) mm and different widths (3.5,4) mm.
- All involved implants were implanted at a minimum distance of 1.5 mm away from the adjacent teeth and a 3mm distance between 2 implants.
- Implants were placed 1mm subcrestally in the subcrestal group and at the bone crest in the crestal group.
- The stability of the implant was immediately measured after the surgery usingwas checked by Osstell (ISQ®) in a quotient scale (ISQ).

- The performed measurement will include 3 different directions: the buccal, mesial, and distal, therefore, therecorded readingswasthe mean of the three readings for each dental implant (21).
- A cover screw was connected to the implant and sutures were taken using 3/0 silk sutures.
- After about 3-4 months period of time from the surgery, an incision was performed directly over the fixed implant and the cover screw was taken out.
- The Implant stability was remeasured by using the Osstell device.

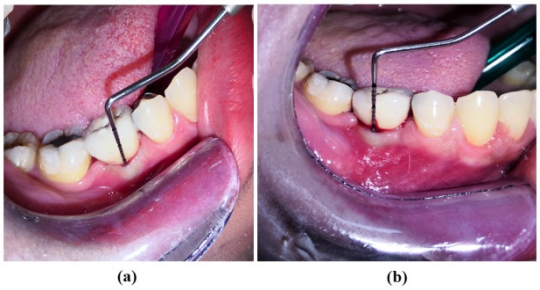
Post-surgical phase (4)

After implant placement and suturing, patients were instructed to follow a soft diet and to avoid chewing at the surgery site until suture removal. They were advised to take the prescribed medications, which include 1 gm tablet of Augmentin (Amoxicillin 875 mg + Clavulanic acid 125 mg: GlaxoSmithKline, UK.) Orally every 12 hours for 7 days. Cataflam 50 mg tablets (Diclofenac. Potassium Novartis, Switzerland) as an analgesic every eight hours and 0.12% chlorhexidine mouthwash (Arab drug company, ADCO) 3 times per day for2 weeks long starting the day after surgery. Sutures were taken out 7 days post-surgery. Sutures were removed after one week and a clinical evaluation of the surgical site was done.

Follow- up phase (Fig 3,4)

(a) (b)

Figure 3: Probing depth (crestal group)



(a) (b)

Figure 4: Probing depth (subcrestal group)

Both clinicaland radiographical follow-ups were done for the patients for about 6 months.

A) Clinical evaluation

- At the permanent prosthesis placement, patients were informed by careful oral-hygiene instructions. The initial peri-implant clinical parameters (plaque index, peri implant probing depth and bleeding index) were taken 1 month (T1) and then after 3 months from loading time (T2) to assess:

1) Modified Plaque index (22)

- It was assessed 1 month after loading (T1) and 3 months after loading (T2) at four aspects around each performed implant (buccal, lingual, mesial and distal) then the assessed mean was calculated.
- It was done by visual inspection using a mouth mirror and a periodontal probe after the abutments air drying.

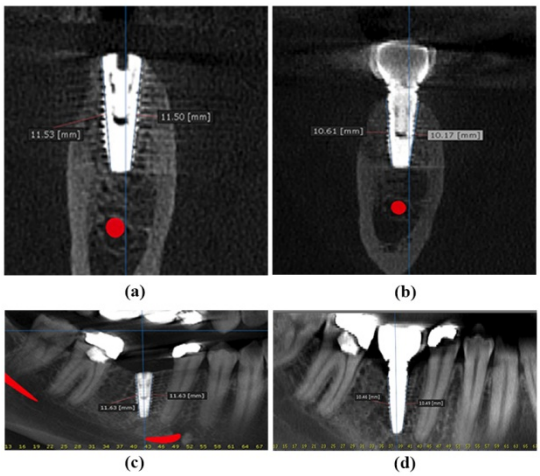
2) Peri-implant probing depth (PPD) (23)

A graduated PDL probe (a manual UNC 15-mm periodontal probe (Hu-Friedy, Chicago, IL, USA) was placed parallel to the implant`slong axis and introduced lightly to the peri-implant sulcus. Measurements were taken around each implant from 6 sites which arethe mesio- buccal, mid-buccal, distobuccal, mesio-palatal, mid-palatal, and disto- palatal 1 month after loading (T1) then 3 months after loading(T2), and the mean will be calculated after that. The probe readingwith 1mm or less depth was registered as 1 mm while the probe reading of more than 1 mm, but less than 2 mmdepth was consideredabout 2 mm.

3) Modified bleeding index (22)

A score of 0-3 was assigned at 4 points (buccal, lingual, mesial, and distal) around eachimplant and the mean was calculated 1 month after loading (T1) and then after 3 months from loading(T2).

B) Radiographic evaluation (Fig 5,6)



(a) (b)

(c) (d)

Figure 5: Radiographic CBL of crestal group

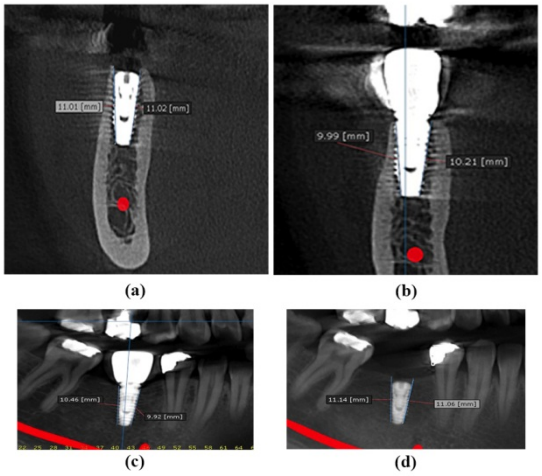


Figure 6: Radiographic CBL of subcrestal group

CBCT was done 3 times: T0 was immediatelypost-surgery, T2 was at loading time (3 months after 1st stage surgery), and T3 was taken after 6 months from 1st stage surgery. The image was analyzed using OnDemand3D™ software CBCT analyzing software. The ruler tool was used to measure the height starting from the bone crest up to the implant apical end to evaluate any marginal bone loss occurring in all aspects (mesial, distal buccal, and lingual)(24).During each follow-up period, thedistance difference from the bone crest to the implant apical end was assessed then the mean was formed for both the mesial and distal and also for the buccal and lingual.

Statistical analysis

Normality was checked using the Shapiro-Wilk test, box plots, and descriptives. Variables were presented using all of the following values: Mean, Standard deviation, Median, Inter Quartile Range (IQR), and Minimum and Maximum values. Comparisonsamong the study groups were done using the Mann-Whitney U test regarding both the plaque and bleeding index and loss in crestal bone while Wilcoxon sign rank was applied for intragroup comparisons. An independentt-test was performed to evaluateboth the implant stability and probing depth differences between the study groups and paired t-test for before and after comparisons. The level of significance was set at a P value of about 0.05. All tests were two-tailed tests. Analysis was performed for the study recorded data using SPSS for windows version 23.

RESULTS

The selected patients' age range was 48.50 years in group I (crestal group) and 48.11 years in group II (subcrestal group). There was one implant failed in group II and two dropouts one in group I and one in group II who didn't respond to their planned follow-up visits. As a result, the total sample was 19 patients (19 implants) 10 for group A and 9 for group B (Table 1).

Plaque index

There was no statistically significant difference among the 2 study groups (P=0.126) including the mean plaque index which was 0.00±0.00 in group I and 0.19 ±0.39 in group II 1 month after loading by (T1). Meanwhile, at (T2) the assessed plaque index was higher in subcrestal group exhibiting a difference that was considered statistically significant between the study groups (p= 0.001). In groupI, the calculated mean was about 0.50±0.00, while in group II it was 0.77±0.19. (Table 2)

Modified bleeding index

At T1 the difference was statistically significant between the 2 study groups (P=0.023). The calculated mean of the bleeding index in group I was about 1.35±0.41 while in group IIwas 1.77±0.36. At T2 the difference was not statistically significant between the 2 groups. (P=0.493) the calculated mean was 2.35±0.53 and 2.22±0.44 in group I and II respectively. (Table 2)

Peri implant probing depth (PPD)

The difference between the two groups at T1 (P=0.117) and T2 (P=0.066) was not significant. At T1 the calculated mean of the probing depth in group I and group II were 3.43±0.08 and 3.68±0.47 respectively. At T2 the mean probing depth was 4.25±0.21 while in group II it was 4.44±0.22. (Table 2)

Implant stability

The difference found between the 2 groups immediately after implant placement (P=0.486) was not statistically significant. The mean implant stability quotient value was recorded as 61.10 ±2.71 in group I while in group II it was 62.44 ±5.25. The difference between the 2 groups (P=0.033) three months after the implant placement was statistically significant. The mean ISQ was 70.55±3.71 in group I while it was higher in group II with a mean of 76.75±7.49. (Table 2)

Table 1: Demographic data of the study sample

		Crestal group (n=10)	Sub-crestal group (n=9)	Test (P value)
Age: Mean (SD)		48.50 (7.01)	48.11 (9.02)	0.106 (0.917)
Gender: n (%)	Males	6 (60%)	5 (55.6%)	0.038 (1.00)
	Female	4 (40%)	4 (44.4%)	

Table 2: Clinical variables between study groups

		Crestal group (n=10)	Sub-crestal group (n=9)	Test (P value)
Plaque Index				
Before	Mean (SD)	0.00 (0.00)	0.19 (0.39)	0.126 **(0.126)**
	Median (IQR)	0.00 (0.00)	0.00 (0.38)	
	Min - Max	0.00 – 0.00	0.00 – 1.00	
After	Mean (SD)	0.50 (0.00)	0.77 (0.19)	3.329 **(0.001*)**
	Median (IQR)	0.50 (0.00)	0.75 (0.38	
	Min - Max	0.50 – 0.50	0.50 – 1.00	
Test (P value)		2.555 **(0.011*)**	3.162 **(0.002*)**	
Bleeding Index				
Before	Mean (SD)	1.35 (0.41)	1.77 (0.36)	2.271 **(0.023*)**
	Median (IQR)	1.50 (0.50)	2.00 (0.50)	
	Min - Max	0.50 – 2.00	1.00 – 2.00	
After	Mean (SD)	2.35 (0.53)	2.22 (0.44)	0.686 **(0.493)**
	Median (IQR)	2.50 (0.75)	2.50 (0.75)	
	Min - Max	1.50 – 3.00	1.50 – 2.50	
Test (P value)		2.970 **(0.003*)**	2.111 **(0.035*)**	
Probing Depth				
Before	Mean (SD)	3.43 (0.08)	3.68 (0.47)	1.653 **(0.117)**
	Median (IQR)	3.50 (0.17)	3.50 (0.75)	
	Min - Max	3.33 – 3.50	3.33 – 4.50	
After	Mean (SD)	4.25 (0.21)	4.44 (0.22)	1.961 **(0.066)**
	Median (IQR)	4.25 (0.50)	4.50 (0.33)	
	Min - Max	4.00 – 4.50	4.00 – 4.67	
Test (P value)		12.943 **(<0.0001*)**	6.252 **(<0.0001*)**	
Stability				
Before	Mean (SD)	61.10 (2.71)	62.44 (5.25)	0.712 (0.486)
	Median (IQR)	60.25 (3.75)	60.50 (5.13)	
	Min - Max	57.50 – 67.00	58.50 – 75.25	
After	Mean (SD)	70.55 (3.71)	76.75 (7.49)	2.324 **(0.033*)**
	Median (IQR)	70.37 (5.56)	76.50 (16.63)	
	Min - Max	64.75 – 76.25	67.25 – 85.75	
Test (P value)		15.823 **(<0.0001*)**	8.523 **(<0.0001*)**	

*Statistically significant at p value<0.05

Crestal bone loss (Buccolingual)

The difference between the 2 study groups regarding the buccolingual CBL at T1 (loading time) (P= 0.870) and T2 (3 months after loading) (P=0.743) wasn`t significant. At (T1) in group I CBL recorded a mean of -0.81±0.34 and -0.80±0.23 in group II. At T2 the calculated mean reachedabout -1.09±0.37 in group I and -1.03±0.25 in group II. (Table 3)

Table 3: Comparison of crestal bone loss between the study groups

		Crestal group (n=10)	Sub-crestal group (n=9)	Test (P value)
Crestal bone loss (Buccolingual)				
Loading (T1)	Mean (SD)	-0.81 (0.34)	-0.80 (0.23)	0.164 **(0.870)**
	Median (IQR)	-0.71 (0.71)	-0.86 (0.32)	
	Min - Max	-1.27 – -0.39	-1.07 – -0.41	
3 months (T2)	Mean (SD)	-1.09 (0.37)	-1.03 (0.25)	0.327 **(0.743)**
	Median (IQR)	-1.05 (0.69)	-1.08 (0.29)	
	Min - Max	-1.55 – -0.56	-1.42 – -0.65	
Test (P value)		2.809 **(0.005*)**	2.670 **(0.008*)**	
Crestal bone loss (Mesiodistal)				
Loading (T1)	Mean (SD)	-1.16 (0.22)	-0.37 (0.38)	2.046 **(0.041*)**
	Median (IQR)	-1.07 (0.41)	-0.47 (0.09)	
	Min – Max	-1.56 – -0.61	-0.55 – 0.64	
3 months (T2)	Mean (SD)	-1.16 (0.22)	-0.79 (0.36)	2.864 **(0.004*)**
	Median (IQR)	-1.07 (0.41)	-0.88 (0.26)	
	Min – Max	-1.56 – -0.61	-1.10 – 0.11	
Test (P value)		2.807 **(0.005*)**	2.680 **(0.007*)**	

*Statistically significant at p value<0.05
Negative sign in CBL means Bone loss

Crestal bone loss (mesiodistally)

The difference between the 2 study groups at T1(P=0.041) and T2 (P=0.004) was statistically significant. At T1 CBL mesiodistally recorded a mean of -0.75±0.30 in group I while forgroup II the calculated mean was -0.37±0.38. At T2 the mean of CBL reached -1.16 ±0.22 in group I and -0.79±0.36 in group II. (Table 3)

DISCUSSION

Although dental implants surpass other traditional restorations in terms of function and aesthetics, their usage could be an absolute or relative risk in

individuals with certain systemically related disorders, such as diabetes mellitus (25). The dental implants' long-term survival and success require the peri-implant tissue stability state and crestal bone preservation. (26).

Regarding bleeding index and peri-implant probing depth, our results agreed with Pellicer *et al*results which disclosed that the differences between the 2 groups were not of statistical significance (20). Our results were inconsistent with the study by Boynueri*et al* who stated that BOP sites were lower in crestal group implants than in subcrestal group, but they used a polished neck implant in their study (27).

BOP is a significant indication of peri-implant health(20). BOP values were high throughout our study (at T1 and T2). At T2 the mean bleeding index reached 2.35 ±0.53 in crestal group and 2.22 ±0.44 in subcrestal group. There is an explanation for that as persistent hyperglycemia stimulates proinflammatory cytokines production such as IL-6 which is producedby the human gingival fibroblasts(28). Also, in T2DM oral hygiene conditions may impact inflammatory markers and consequently cause periimplantitis(29). Our results agreed withthe meta-analysis by Lagunov*et al* who indicated a significant rise in BOP and PD when comparing the controlled T2DM (HbA1C 6.1%-8%) to healthy individuals in 4 studies (30).

Different probing depths have been recorded in numerous studies. Duarte et al recorded a mean PPD of 3.7 ± 0.8(31). Also, Pellicer *et al* recorded a mean of 2.97±0.90 mm and 3.06±0.88 mm in crestal and subcrestal groups respectively(20).

The mean of PPD in our study was 4.25 mm ±0.21 in crestal group and 4.44 mm ±0.22 in subcrestal group. Both of these values correspond to normal peri-implant tissues(32).

Among the subcrestal group, the probing depth was higher but with no significant difference. This outcome might be ascribed to Pellicer *et al* who claimed an association that reveals the fact that the deeper the implant location, the deeper the pocket depth would be(12).

Although plaque index was higher in subcrestal group, it was minimal throughout the study as it didn't exceed score 1 (22) (Mombelli score) as the mean was 0.50±0.00 in crestal group and was 0.77±0.19 in subcrestal group 3 months after loading. The cause of the significant difference between the 2 groups may be related to deeper implant placement which leads to subgingival crown margin which makes hygiene control problematic as Saleh et al stated in their study(33). Also, it may be because of noncompliance with a proper technique of tooth brushing or following oral hygiene measures regularly (34).

Regarding the crestal bone loss andstability, at T2 our study showed that the mesiodistal CBL was significantly lower in the subcrestal group P= (0.004) while the difference between both groups in buccolingual CBL P= (0.743) was not statistically significant. In addition, ISQ values were higher with significance in the subcrestal group compared to crestal group (P=0.033).

The majority of studies have used conventional PA radiographs with paralleling techniques to assess mesiodistal (interproximal) marginal or crestal bone loss (35). Only very few studies examined the buccal and lingual aspects.

To our knowledge, there is only one study by Koutouzis*et al* who assessed the marginal bone level on the buccal and lingual aspect in relation to implant placement depth using CBCT(36). In our study, we evaluated mesiodistal as well as buccolingual CBL using CBCT. According to the mesiodistal CBL, our results showed that subcrestal implant placement exhibited significantly less CBL compared to crestal group at T1 P= (0.041) and T2 P= (0.004). Our results have coincidedwith madani *et al* who stated in their study that platform-switched implants inserted subcrestally within 1 mm and 1.99 mm demonstrated that crestal bone loss occurs but to the least extent and that they were not related to any implant radiographical exposure over time(11). However, Ercoli*et al* found that there is no difference in mesio distal CBL between crestal and subcrestal implants although subcrestal placement decreased the odds of thread exposure with time (37).

Bucco lingual CBL differences displayed no significance between crestal and subcrestal groups at T1 P= (0.870) and T2 P= (0.743) respectively while there is less chance for the subcrestal group to be exposed at the alveolar crest (mean = -1.03 mm) compared to crestal group.

Although buccolingual CBL was proved to be non-significant, exposed rough implant surfaces in crestal group might cause difficulties in peri-implant health as the visible roughsurface of an implant migh provide a microenvironment preferable to bacterial contamination as a result of infiltration via the peri-implant sulcus. When implants are inserted at the crestal level, early bone remodeling exposes the implant's rough surface. This does not occur when implants are placed at subcrestal level because the starting point of the bone is already above the platform of the implant and its rough surface is trapped inside the peri-implant defect formed as a result of the drillingstep (20).

Our results coincide with koutouzis et al who found that the percentages of buccolingual surfaces within bone is considerably greater in subcrestal group than epicrestal group(37).

The greater values in subcrestal group may be due to less mesiodistal CBL compared to crestal group, and all implant threads were within bone even if there is no difference between both groups in buccolingual CBL (mean=-1.03mm)

Our result agreed with a study done by Guirado*et al* who stated that after 8 weeks, ISQ values were higher in subcrestal group 71.5 ± 0.67 compared with the crestal group 69.5 ± 0.12(38). While Romanos*et al* reported a non-significant difference for the Periotest values between crestal and subcrestal implants(39).

CONCLUSIONS

From our study results, we can conclude the fact that subcrestal implant placement is associated with less mesiodistal crestal bone loss, less thread exposure, and better stability than crestal implant placement so, placing an implant at a subcrestal level may be favorable to decrease the probability of implant thread exposure, provided that meticulous oral hygiene care and regular periodic checks are kept to maintain both peri-implant soft tissue and bone health in controlled T2DM.

CONFLICT OF INTEREST

The authors reveal that they have not experienced any financial or personal conflicts of interest.

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REFERENCES

1. Kulkarni C, Roshani Thakur D, Jangid M, Hendre P, Sohani A, Kulkarni K. Evaluation of success rate and incidence of periimplantitis in dental implants placed in a post graduate dental institute: a retrospective study. World JAdvSciRes. 2019;2:24-32.
2. Albrektsson T, Zarb G, Worthington P, Eriksson AR. The long-term efficacy of currently used dental implants: a review and proposed criteria of success. IntJ Oral Maxillofac Implants. 1986;1:11-25.
3. Gu Y, Shi J, Zhuang L, Qiao S, Xu Y, Lai H. Esthetic outcome and alterations of soft tissue around single implant crowns: a 2-year prospective study. Clin Oral Implants Res. 2015;26:909-14.
4. Oh TJ, Yoon J, Misch CE, Wang HL. The causes of early implant bone loss: myth or science? J Periodontol. 2002 Mar;73(3):322-33.
5. Mellado-Valero A, FerrerGarcía JC, Herrera Ballester A, Labaig Rueda C. Effects of diabetes on the osseointegration of dental implants. Med Oral Patol Oral Cir Bucal. 2007;12:E38-43.
6. Vissink A, Spijkervet FKL, Raghoebar GM. The medically compromised patient: Are dental implants a feasible option? Oral Dis. 2018;24:253-60.
7. Javed F, Romanos GE. Impact of diabetes mellitus and glycemic control on the osseointegration of dental implants: a systematic literature review. J Periodontol. 2009;80:1719-30.
8. Mostafa SA, Davies MJ, Srinivasan BT, Carey ME, Webb D, Khunti K. Should glycatedhaemoglobin (HbA1c) be used to detect people with type 2 diabetes mellitus and impaired glucose regulation? Postgrad Med J. 2010;86:656-62.
9. Gatti C, Gatti F, Silvestri M, Mintrone F, Rossi R, Tridondani G, et al. A Prospective Multicenter Study on Radiographic Crestal Bone Changes Around Dental Implants Placed at Crestal or Subcrestal Level: One-Year Findings. Int J Oral Maxillofac Implants. 2018;33:913-8.
10. Welander M, Abrahamsson I, Berglundh T. Placement of two-part implants in sites with different buccal and lingual bone heights. J Periodontol. 2009;80:324-9.
11. Madani E, Smeets R, Freiwald E, Sanj MS, Jung O, Grubeanu D, et al. Impact of different placement depths on the crestal bone level of immediate versus delayed placed platform-switched implants. J Craniomaxillofac Surg. 2018 Jul;46:1139-46.
12. Pellicer-Chover H, Peñarrocha-Diago M, Peñarrocha-Oltra D, Gomar-Vercher S, Agustín-Panadero R, Peñarrocha-Diago M. Impact of crestal and subcrestal implant placement in peri-implant bone: A prospective comparative study. Med Oral Patol Oral Cir Bucal. 2016;21:e103-10.
13. Hermann JS, Buser D, Schenk RK, Cochran DL. Crestal bone changes around titanium implants. A histometric evaluation of unloaded non-submerged and submerged implants in the canine mandible.J Periodontol. 2000;71:1412-24.
14. Fenner N, Hämmerle CH, Sailer I, Jung RE. Long-term clinical, technical, and esthetic outcomes of all-ceramic vs. titanium abutments on implant supporting single-tooth reconstructions after at least 5 years. Clin Oral Implants Res. 2016;27:716-23.
15. Abrahamsson I, Berglundh T, Moon I, Lindhe J. Peri-implant tissues at submerged and non-submerged titanium implants. J ClinPeriodontol. 1999;26:600-7.

16. Donovan R, Fetner A, Koutouzis T, Lundgren T. Crestal bone changes around implants with reduced abutment diameter placed non-submerged and at subcrestal positions: A 1-year radiographic evaluation. J Periodontol. 2010;81:428-34.
17. Nisapakultorn K, Suphanantachat S, Silkosessak O, Rattanamongkolgul S. Factors affecting soft tissue level around anterior maxillary single-tooth implants. Clin Oral Implants Res. 2010;21:662-70.
18. Consort. Welcome to the CONSORT Website. Consort. Available from: http://www.consort-statement.org/ [Accessed on: Jan, 2022]
19. World Medical Association. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. JAMA. 2013;310:2191-4.
20. Pellicer-Chover H, Peñarrocha-Diago M, Aloy-Prosper A, Canullo L, Peñarrocha-Diago M, Peñarrocha-Oltra D. Does Apico-Coronal Implant Position Influence Peri-Implant Marginal Bone Loss? A 36-Month Follow-Up Randomized Clinical Trial. J Oral Maxillofac Surg. 2019;77:515-27.
21. Balleri P, Cozzolino A, Ghelli L, Momicchioli G, Varriale A. Stability measurements of osseointegrated implants using Osstell in partially edentulous jaws after 1 year of loading: a pilot study. Clin Implant Dent Relat Res. 2002;4:128-32.
22. Mombelli A, van Oosten MA, Schurch E, Land NP. The microbiota associated with successful or failing osseointegrated titanium implants. Oral MicrobiolImmunol. 1987;2:145-51.
23. Giannopoulou C, Bernard JP, Buser D, Carrel A, Belser UC. Effect of intracrevicular restoration margins on peri-implant health: clinical, biochemical, and microbiologic findings around esthetic implants up to 9 years. Int J Oral Maxillofac Implants. 2003;18:173-81.
24. Patil PG, Nimbalkar-Patil S. A radiographic measurement technique for crest bone changes related to dental implants. J Prosthet Dent. 2015;113:350-1.
25. Fiorellini JP, Chen PK, Nevins M, Nevins ML. A retrospective study of dental implants in diabetic patients.Int J Periodontics Restorative Dent. 2000;20:366-73.
26. Alasqah MN, Alrabiah M, Al-Aali KA, Mokeem SA, Binmahfooz AM, ArRejaie AS, et al. Peri-implant soft tissue status and crestal bone levels around adjacent implants placed in patients with and without type-2 diabetes mellitus: 6 years follow-up results. Clin Implant Dent Relat Res. 2018;20:562-8.
27. Boynueğri AD, Yalim M, Nemli SK, Ergüder BI, Gökalp P. Effect of different localizations of microgap on clinical parameters and inflammatory cytokines in peri-implant crevicular fluid: a prospective comparative study. Clin Oral Investig. 2012;16:353-61.
28. Chiu H, Fu MM, Yang T, Fu E, Chiang C, Tu H, et al. Effect of high glucose, Porphyromonasgingivalis lipopolysaccharide and advanced glycation end-products on production of interleukin-6/-8 by gingival fibroblasts. J Periodontal Res. 2017;52:268-76.
29. MacDonald LL, Aylward N, Sellers E, Sloshower S, Dean HJ. Development of an interprofessional diabetes and oral hygiene education program for youth with type 2 diabetes mellitus.Can J Diabetes. 2012;36:327-31.
30. Lagunov VL, Sun J, George R. Evaluation of biologic implant success parameters in type 2 diabetic glycemic control patients versus health patients: A meta-analysis. J InvestigClin Dent. 2019;10:e12478.
31. Duarte PM, de Mendonça AC, Máximo MBB, Santos VR, Bastos MF, Nociti FH. Effect of anti-infective mechanical therapy on clinical parameters and cytokine levels in human peri-implant diseases.J Periodontol. 2009;80:234-43.
32. Karoussis IK, Salvi GE, Heitz-Mayfield LJA, Brägger U, Hämmerle CHF, Lang NP. Long-term implant prognosis in patients with and without a history of chronic periodontitis: a 10-year prospective cohort study of the ITI® Dental Implant System. Clin Oral Implants Res. 2003;14:329-39.
33. Saleh MHA, Ravidà A, Suárez-López Del Amo F, Lin GH, Asa'ad F, Wang HL. The effect of implant-abutment junction position on crestal bone loss: A systematic review and meta-analysis. Clin Implant Dent Relat Res. 2018;20:617-33.
34. Sreenivasan PK, Prasad KVV, Javali SB. Oral health practices and prevalence of dental plaque and gingivitis among Indian adults. Clin Exp Dent Res. 2016;2:6-17.

35. Veljanovski D, Atanasovska-Stojanovska A, Pivkova-Veljanovska A, Mijiritsky E, Bollen C. The Vertical Soft Tissue Thickness and Subcrestal Implant Placement as Factors for Peri-implant Crestal bone Stability. Open Access Maced J Med Sci. 2021;9:257-63.
36. Koutouzis T, Neiva R, Nair M, Nonhoff J, Lundgren T. Cone beam computed tomographic evaluation of implants with platform-switched Morse taper connection with the implant-abutment interface at different levels in relation to the alveolar crest. Int J Oral Maxillofac Implants. 2014;29:1157-63.
37. Ercoli C, Jammal G, Buyers M, Tsigarida AA, Chochlidakis KM, Feng C, et al. Influence of apico-coronal implant placement on post-surgical crestal bone loss in humans. J Periodontol. 2017;88:762-70.
38. Calvo-Guirado JL, Delgado Ruiz RA, Ramírez-Fernández MP, Abboud M, Janjic B, Maté Sánchez de Val JE. Retracted: Histological and histomorphometric analyses of narrow implants, crestal and subcrestally placed in severe alveolar atrophy: a study in foxhound dogs. Clin Oral Implants Res. 2016;27:497-504.
39. Romanos GE, Aydin E, Gaertner K, Nentwig G. Long-term results after subcrestal or crestal placement of delayed loaded implants. Clin Implant Dent Relat Res. 2015;17:133-41.